



Australian Government
Biosecurity Australia

4 May 2007

VETERINARY CERTIFICATE FOR CAMELID EMBRYOS FROM THE UNITED STATES OF AMERICA (USA)

Permission to import may also be required from the Australian Government Department of the Environment and Water Resources on telephone 02 6274 1900, facsimile 02 6274 1921 or by email: wildlifetrade@dew.gov.au.

These conditions apply only to the South American camelids – *Lama glama* (llama), *Lama pacos* (alpaca) and *Lama guanaco* (guanaco).

In these conditions

- the *donor* refers to both the female and male donor of germplasm. They were kept isolated from other animals not of equivalent animal health status for 30 days prior to and during the collection period.
 - the minimum and maximum collection period for a donor for each consignment is one and four months respectively. Only embryos collected during this period are eligible for export.
1. The USA is recognised by the World Organisation for Animal Health (OIE) as a foot and mouth disease (FMD) free country where vaccination is not practised and meets the relevant OIE Code Article definitions for country freedom from rinderpest and Rift Valley fever.
 2. During the 12 months prior to, and during, the collection period, infection with *Trypanosoma evansi* (surra) was not reported in the USA.
 3. Immediately prior to, and during, the collection period, each donor was living in the USA, free from quarantine restrictions and showed no clinical evidence of disease.
 4. All blood and tissue tests for disease were carried out at a laboratory approved by US Department of Agriculture (USDA) to perform the test required for that disease. Dates of collection for tests and types of diagnostic tests were recorded on the Veterinary Certificate. Where diagnostic tests are not specified, only those tests recognised by USDA/Animal and Plant Health Inspection Service (APHIS) for the disease were used.
 5. **Bovine brucellosis**
Immediately prior to, and during, the collection period, each donor either:
 - . was part of a herd in a Class Free State* or area,
 - . was part of a herd in which no case of bovine brucellosis occurred during the past 5 years, and
 - . gave a negative result to a complement fixation test (CFT) for bovine brucellosis during the collection period for this consignment.or
 - . was part of a herd in a Class A State* or area,

- . was part of a herd in which no case of bovine brucellosis occurred during the past 5 years, and
- . gave negative results to the CFT for bovine brucellosis on each of two occasions, 30 days apart, during the collection period for this consignment.

[The veterinary certificate must indicate the option that applies]

*[*USDA/APHIS classification]*

6. **Bovine tuberculosis**

Immediately prior to, and during, the collection period, each donor either:

- . was part of a herd in an accredited free US State or zone,
- . was part of a herd in which no case of bovine tuberculosis occurred during the past 5 years, and
- . was tested for bovine tuberculosis by an approved single intradermal tuberculin test at the axillary site (using 0.1ml of USDA tuberculin PPD), with negative results (negative being no swelling or a swelling not greater than 2mm at the site of injection 72 hours after injection), within 14 days prior to the collection period for this consignment but more than 90 days after any previous tuberculin test.

or

- . was part of a herd in a modified accredited State or zone,
- . was part of a herd in which no case of bovine tuberculosis occurred during the past 5 years, and
- . was tested for bovine tuberculosis by an approved single intradermal tuberculin test at the axillary site (using 0.1ml of USDA tuberculin PPD), with negative results (negative being no swelling or a swelling not greater than 2mm at the site of injection 72 hours after injection), twice, the first being within 1 year prior to the collection period and the second being within 14 days prior to the collection period for this consignment but more than 90 days after the previous tuberculin test.

[Note: Reactors to the test, and all in-contact animals, were rejected for export.]

7. **Vesicular stomatitis**

During the collection period, vesicular stomatitis was not reported within 15 kilometres of the collection premises.

8. **Bluetongue**

Each donor was either

- . kept in a bluetongue virus free, or seasonally free, zone for at least the 60 days before, and during, the collection period,

or

- . subjected to a serological test to detect antibody to the bluetongue virus group, such as the bluetongue competition enzyme-linked immunosorbent assay (ELISA) or the bluetongue agar gel immunodiffusion (AGID) test, between 28 and 60 days after the final collection for this consignment, with negative results,

or

- . subjected to a bluetongue virus isolation test or polymerase chain reaction (PCR) test on a blood sample taken on the day(s) of collection during the collection period, with negative results.

9. **Epizootic haemorrhagic disease (EHD)**

Each donor was either

- . kept in an EHD virus free, or seasonally free, zone for at least the 60 days before, and during, the collection period,

or

- . subjected to a serological test to detect antibody to the EHD virus group, such as an AGID test or the virus neutralisation test (VNT), between 28 and 60 days after the final collection for this consignment, with negative results,

or

- . subjected to an EHD virus isolation test or PCR test on a blood sample taken on the day(s) of collection, with negative results.

10. **Bovine pestivirus**

Blood samples, drawn from each donor before the collection period, gave negative results to one of the following tests for bovine pestivirus, either

- . an antigen-capture ELISA on peripheral blood leucocytes

or

- . a virus isolation test on blood or serum

or

- . PCR on peripheral blood leucocytes.

11. **Johne's disease**

During the collection period, donors showed no clinical signs of Johne's disease.

12. **Collection, handling and storage**

The embryos were collected, handled and stored in accordance with Appendix 3.3.1. of the OIE Terrestrial Animal Health Code.

ROBYN MARTIN
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Animal Biosecurity